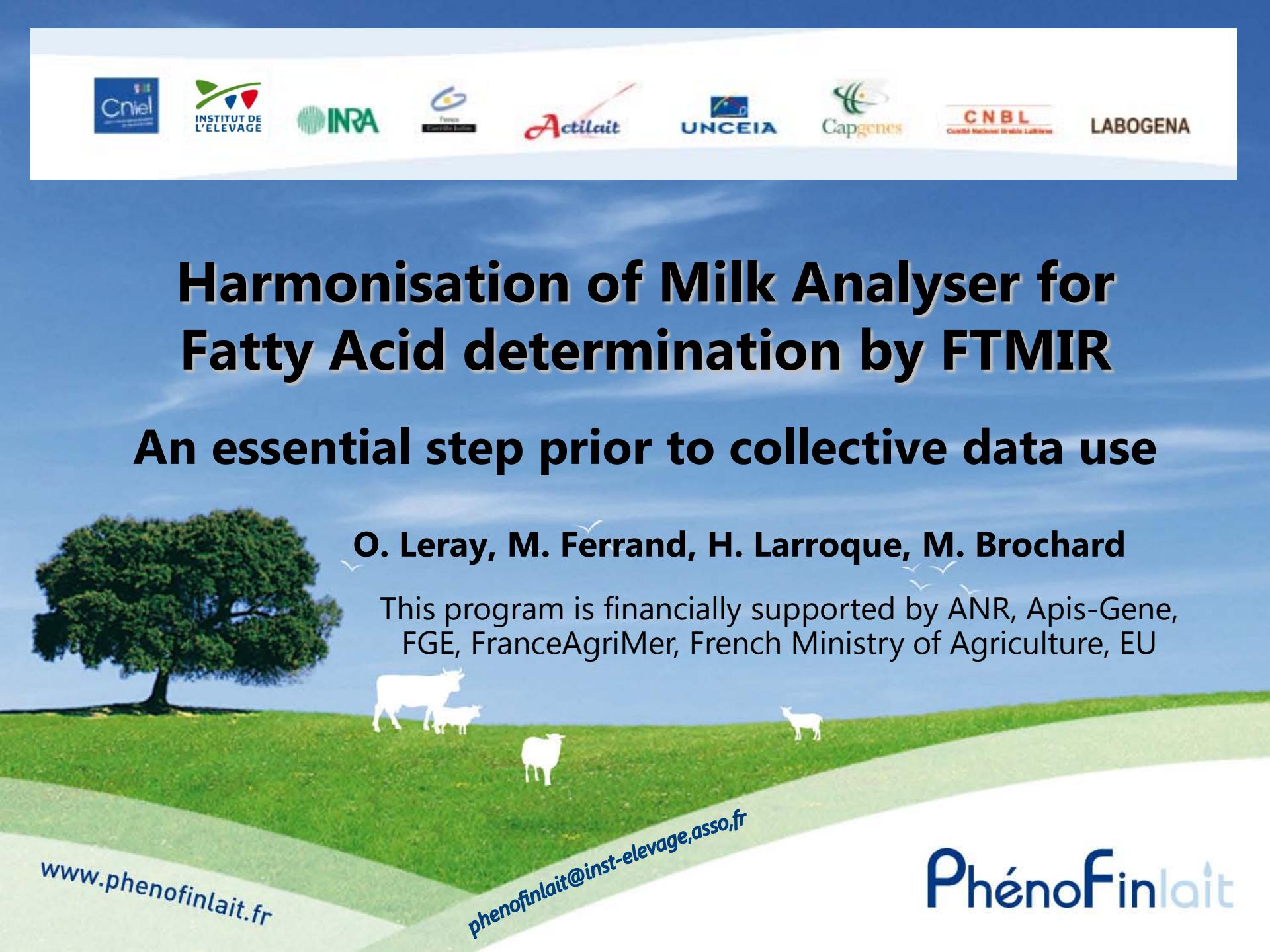


Harmonisation of Milk Analyser for Fatty Acid determination by FT-MIR

An essential step prior to collective data use



O. Leray, M. Ferrand, H. Larroque, M. Brochard

This program is financially supported by ANR, Apis-Gene, FGE, FranceAgriMer, French Ministry of Agriculture, EU



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Un programme R&D pour les filières laitières de demain

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Summary of Phénofinlait

- Phenotyping dairy animals for fine milk composition (Fatty Acids FA & Individual Protein IP)
- Fatty acid profiling of individual milk (cow, sheep, goat) by mid-infrared spectrophotometry (FTMIR)
- Establishment of calibration equation sets (cow, sheep, goat)
- Application to about 20 000 spectrum data collected on 14 months with 13 milk FTMIR analysers in 9 laboratories and stored in a spectral data base
- Parallel survey of feeding, parentage, DNA mapping, etc, to establish suitable relations for further data analysis.
- Establishment of diagnosis tools and levers for farmers to monitor and orient milk fine composition.



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Players and Leaders

Dairy Industry Interprofession (CNIEL)

Dairy Genetic Interprofession (FGE) :

- Artificial Insemination Organisations
(UNCEIA, ANIO, CapGène)

- DHIA Federation (FCEL, CNBL)

(MRO : 26 departments)

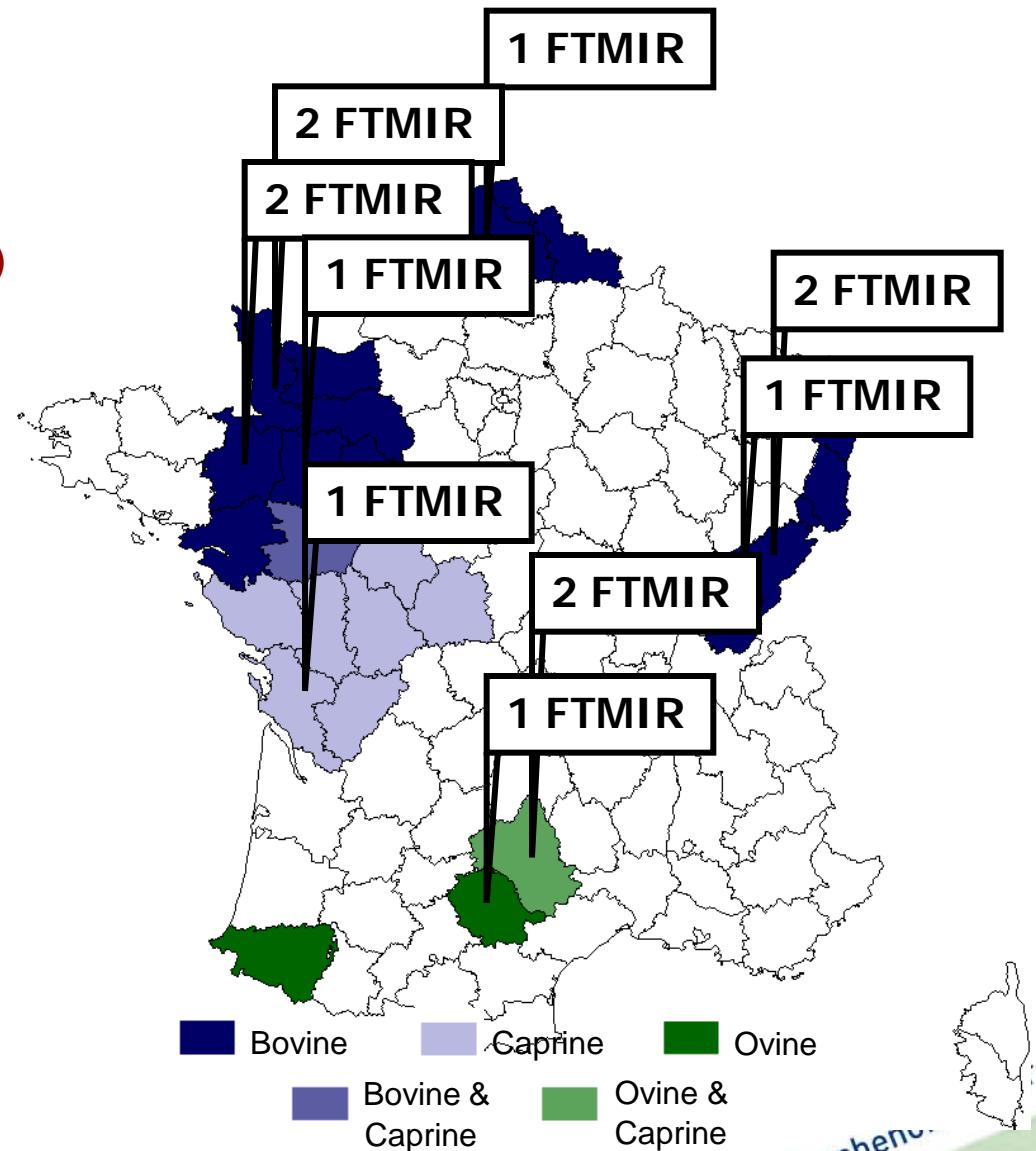
- Actilait & DHI laboratoires

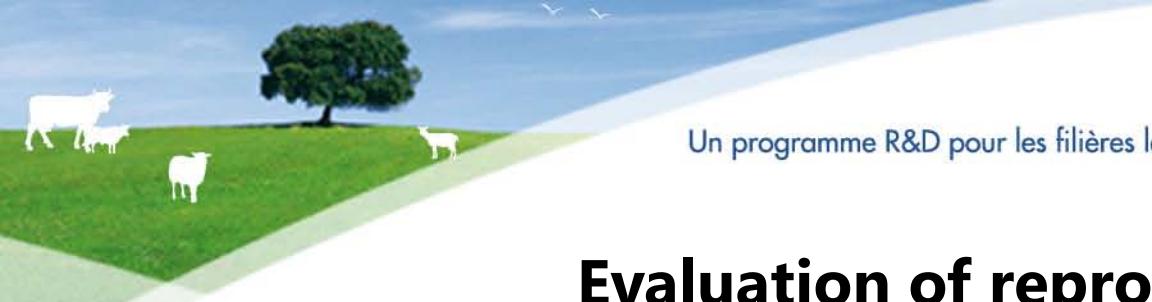
(9 labs, 13 FTMIR)

- INRA (4 labs, 4 exp. units, 2 dpts)

- Institut de l'Elevage (4 teams, 3 dpts,)

... and about 20 000 females
of 1500 herds





Un programme R&D pour les filières laitières de demain

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Evaluation of reproducibility between FTMIR milk analysers

- Interlaboratory study (October 2008):
- 8 laboratories
- 12 Milkoscan FT6000 (Foss, DK)
- 15 individual bovine milk samples
- 13 calibration samples (mid-range MIR RM set, Actilait)
- Analysed in triplicates in spectrum extraction mode (calibration) with requested prior spectrum standardisation
- Application of Phénofinlait equation sets on spectral data for FA prediction



Outcomes

- Heterogeneous distribution of biases between analysers
- High relative biases and bias ranges resulting in high reproducibility values R depending on FA equation
- Biases with natural individual samples and biases with calibration samples are tightly correlated hence calibration is possible.



Evaluation of correction modes

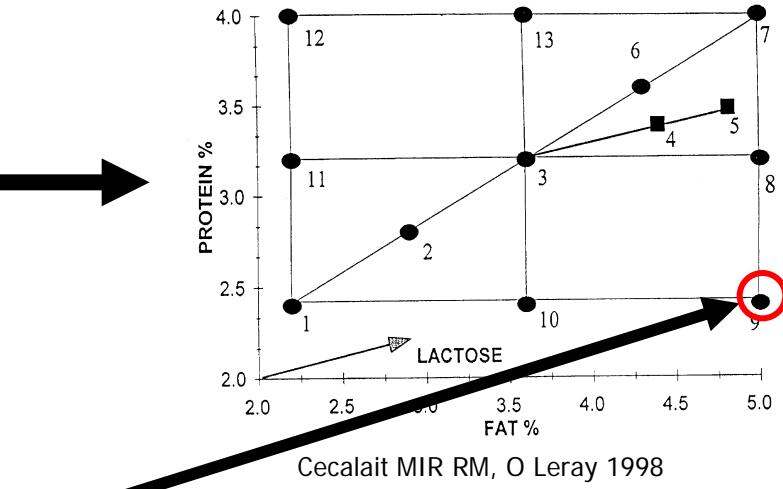
Predictions :

- Linear regression thr. 13 calibration samples



Predictions and absorbances :

- Multiplicative correction thr. 1 sample (x)
- Additive correction thr. 1 sample (+)



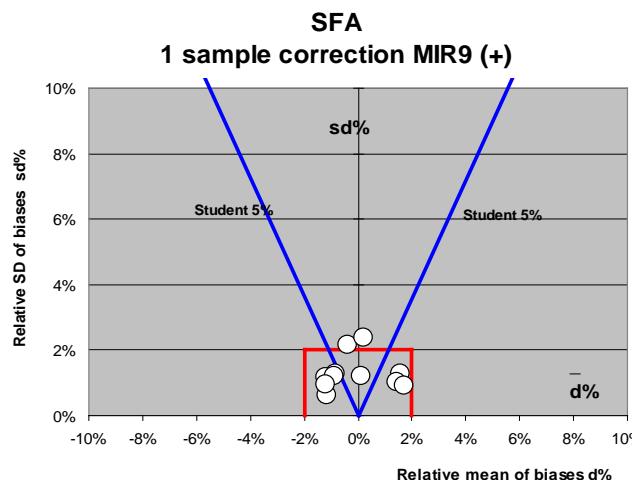
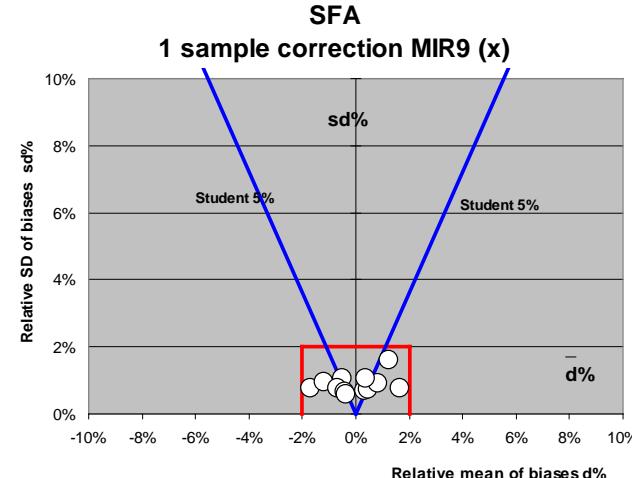
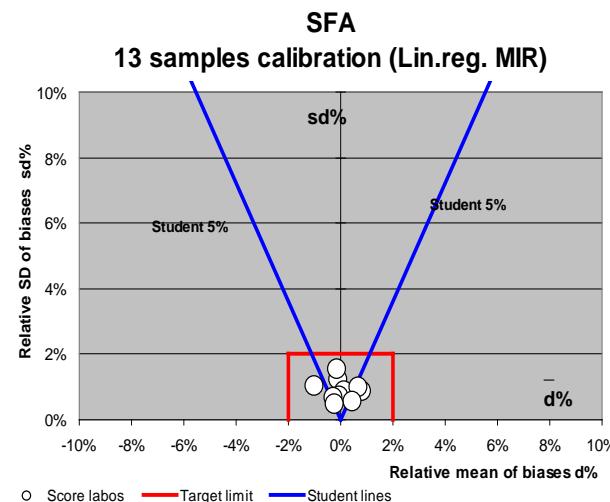
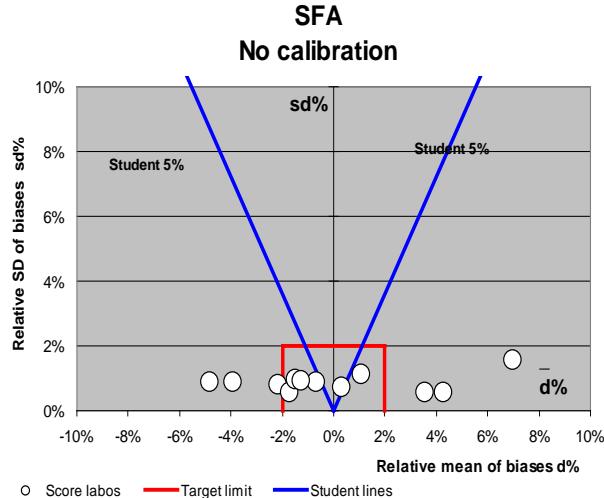
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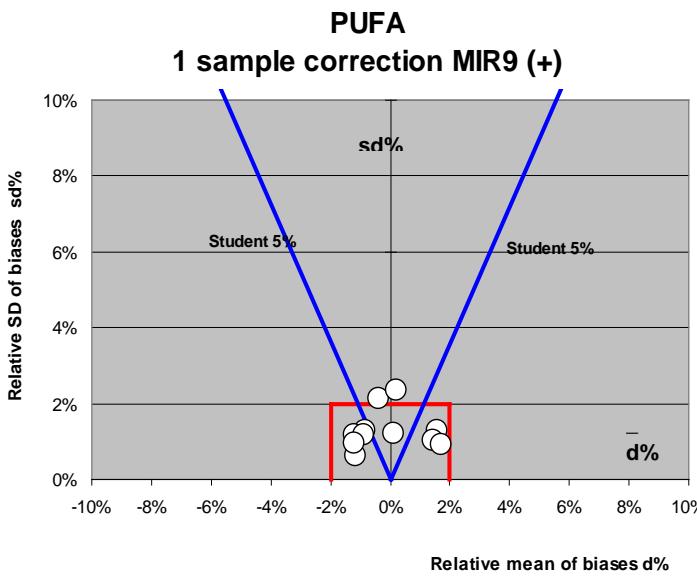
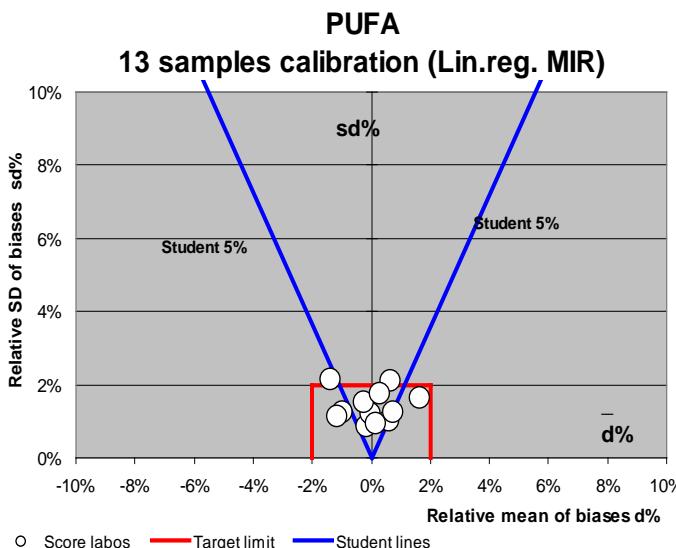
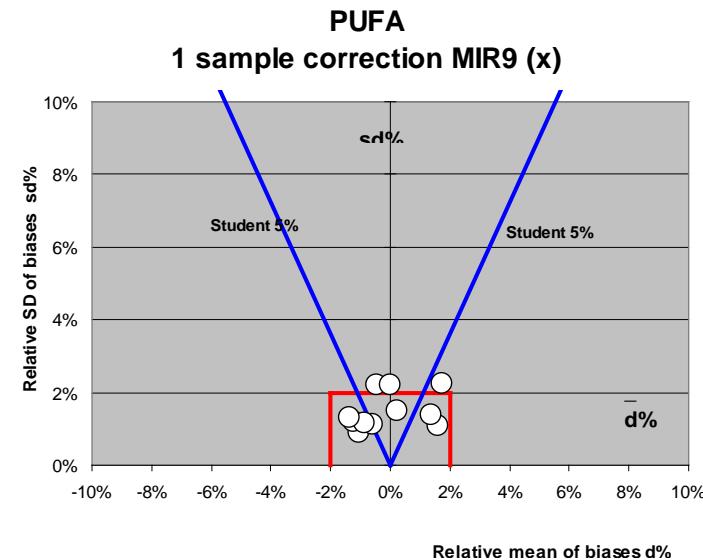
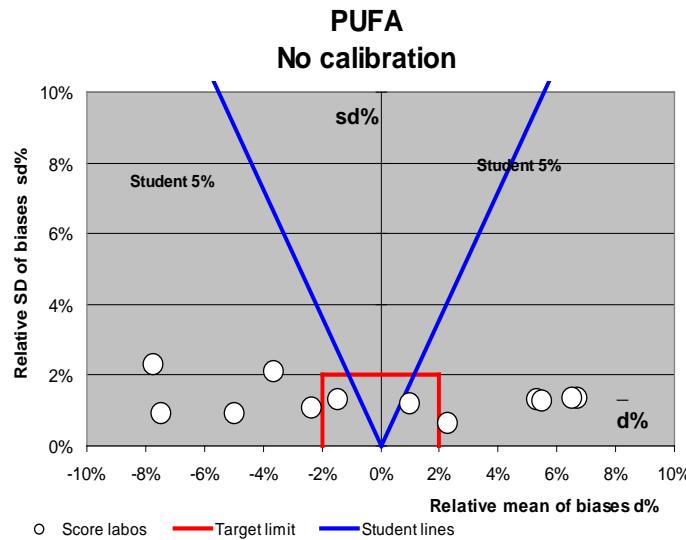
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Correction of predictions – Saturated Fatty Acids





Correction of predictions – Polyunsaturated Fatty Acids



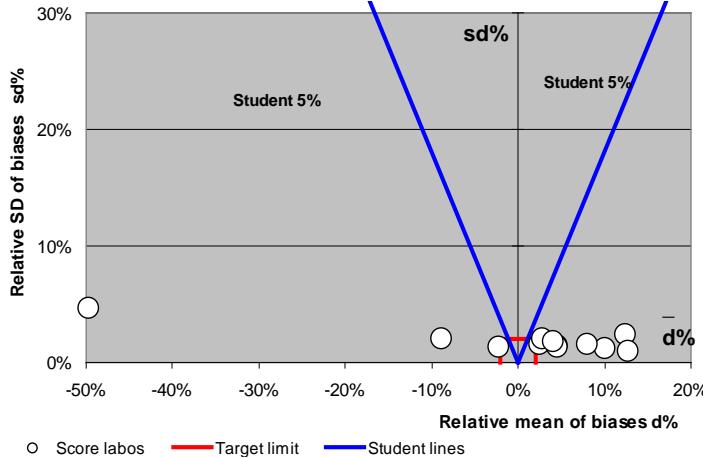


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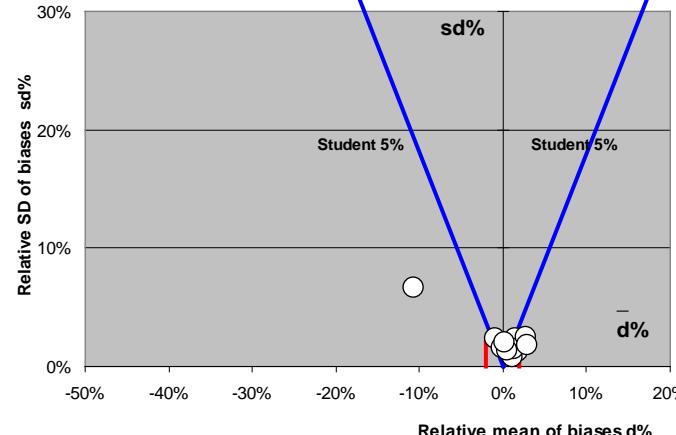
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Correction of predictions – Omega 6

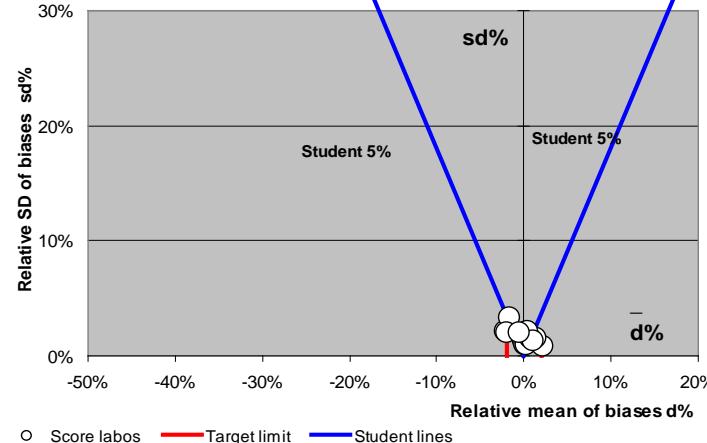
**Omega 6
No calibration**



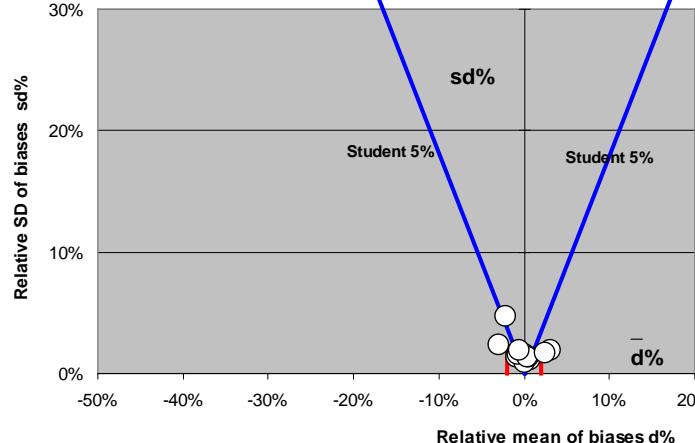
**Omega 6
1 sample correction MIR9 (x)**



**Omega 6
13 samples calibration (Lin.reg. MIR)**



**Omega 6
1 sample correction MIR9 (+)**

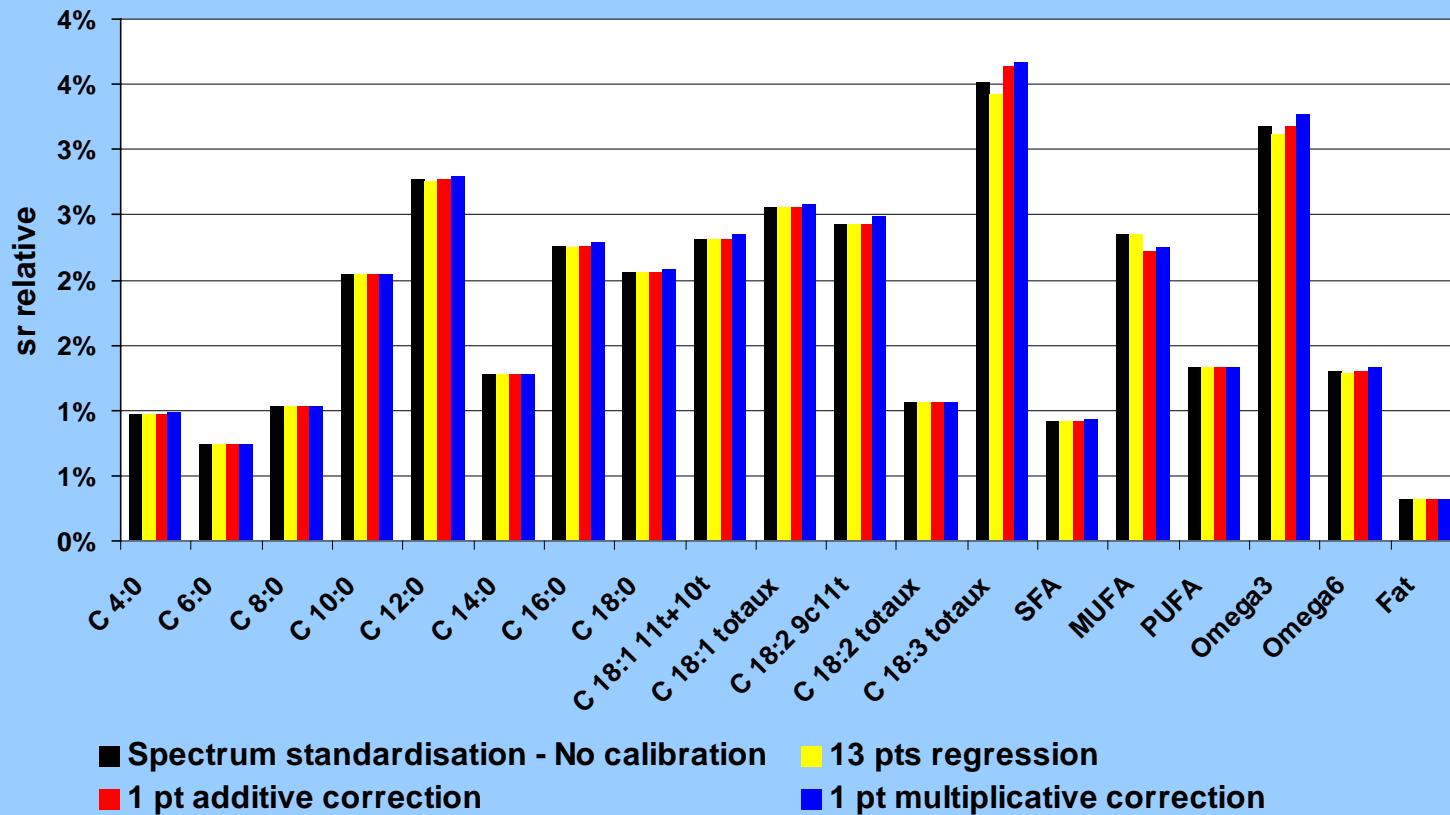




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FTMIR Fatty Acids Determination in Milk
Relative repeatability standard deviation sr rel
Influence of calibration / correction mode

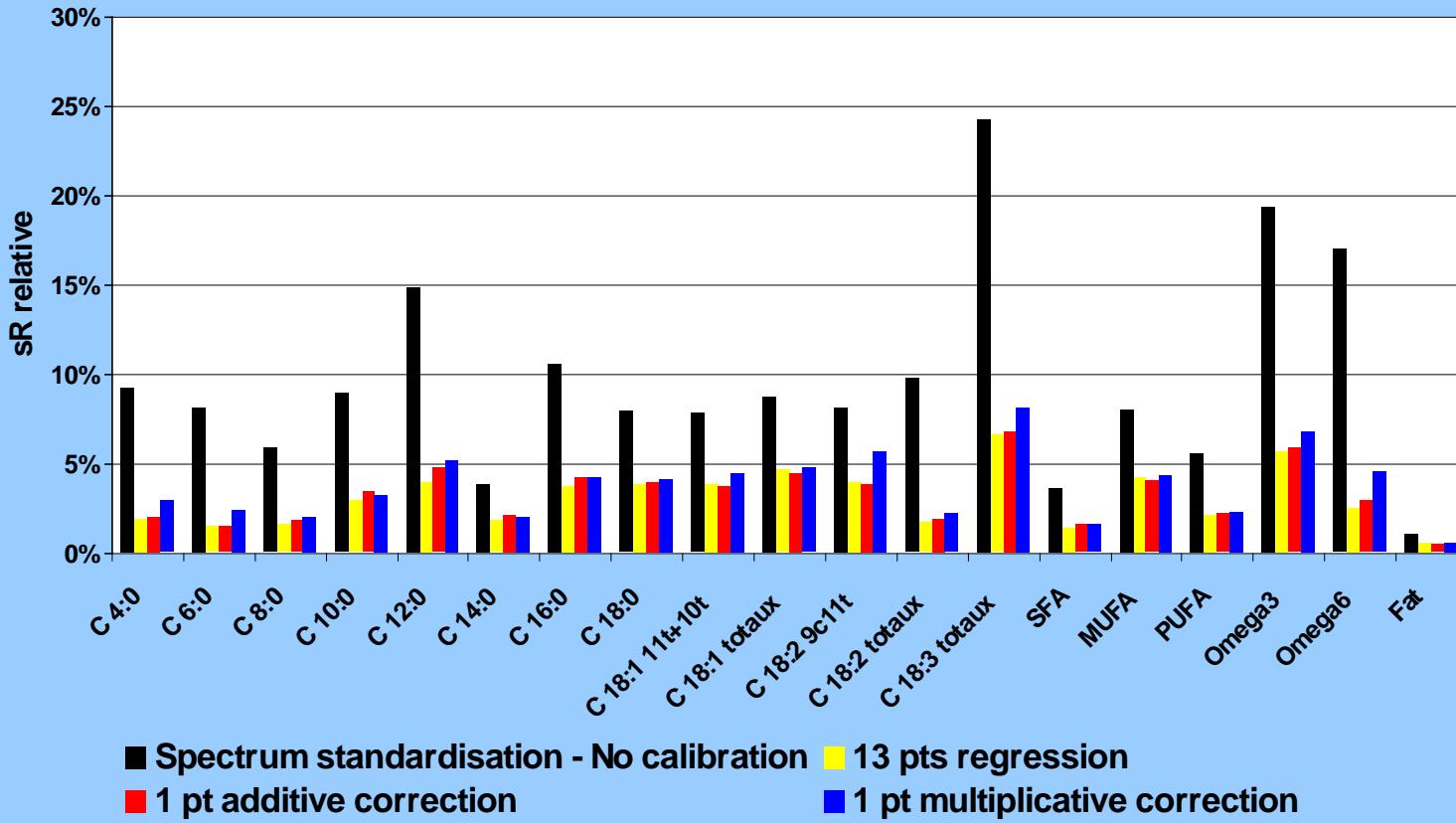




Un programme R&D pour les filières laitières de demain

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FTMIR Fatty Acids Determination in Milk
Relative reproducibility standard deviation sR rel
Influence of calibration / correction mode



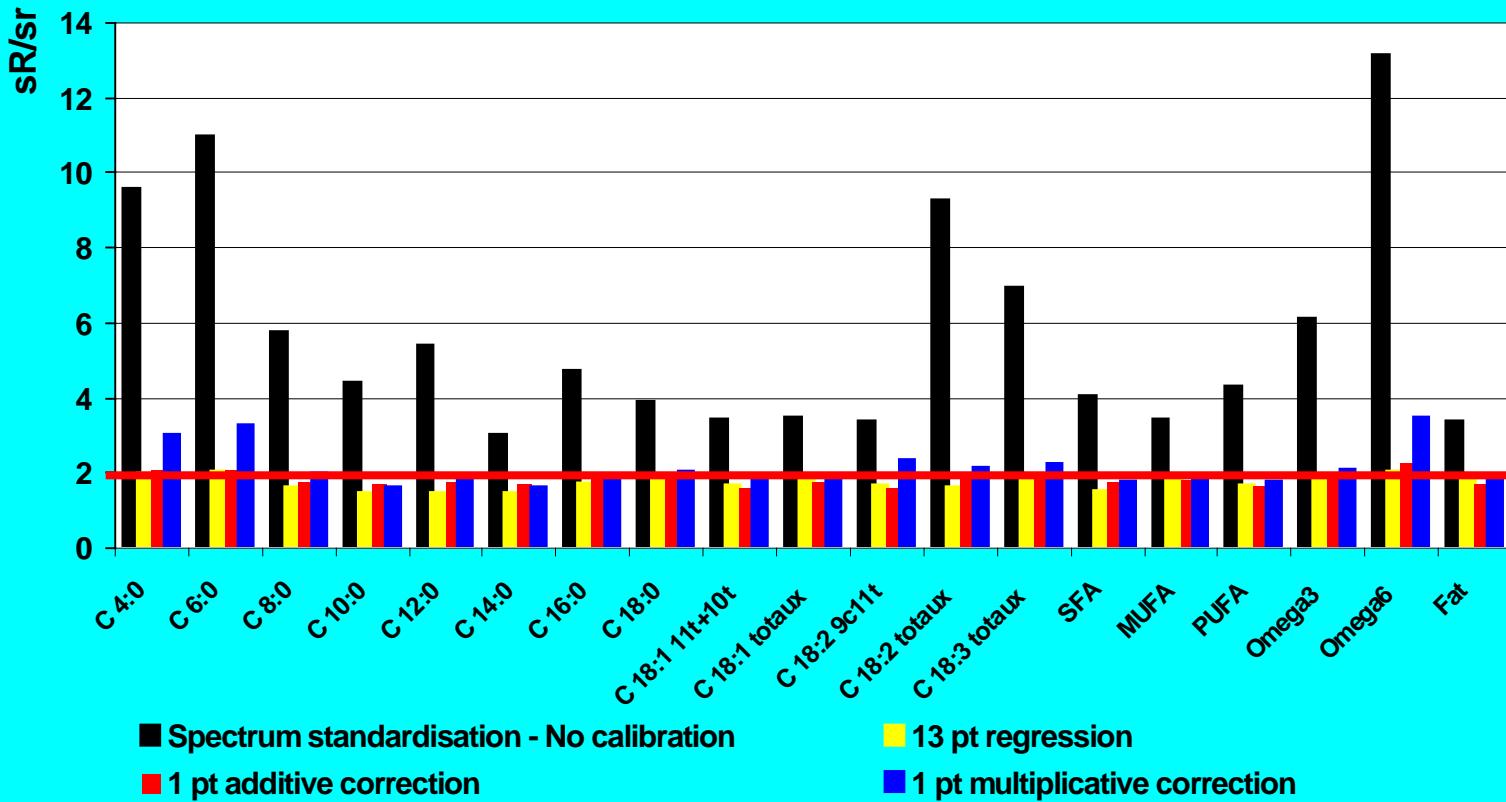


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FTMIR Fatty Acids Determination in Milk

- Precision ratio R/r depending on the calibration / correction mode -





Un programme R&D pour les filières laitières de demain

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Repeatability and reproducibility of FTMIR analysers

| Fatty acids | Mean level g/kg | Spectrum standardisation | | | | Centralised calibration | | | |
|----------------|--------------------|--------------------------|-------|------|-------|-------------------------|-------|------|-------|
| | | r | r rel | R | R rel | r | r rel | R | R rel |
| C 4:0 | 1,84 | 0,05 | 2,7% | 0,48 | 26,1% | 0,05 | 2,7% | 0,10 | 5,3% |
| C 6:0 | 1,05 | 0,02 | 2,1% | 0,24 | 22,7% | 0,02 | 2,1% | 0,04 | 4,2% |
| C 8:0 | 0,59 | 0,02 | 2,9% | 0,10 | 16,7% | 0,02 | 2,9% | 0,03 | 4,6% |
| C 10:0 | 1,23 | 0,07 | 5,7% | 0,31 | 25,1% | 0,07 | 5,7% | 0,10 | 8,3% |
| C 12:0 | 1,36 | 0,11 | 7,8% | 0,57 | 41,8% | 0,11 | 7,7% | 0,15 | 11,0% |
| C 14:0 | 4,57 | 0,16 | 3,6% | 0,50 | 10,8% | 0,16 | 3,6% | 0,24 | 5,2% |
| C 16:0 | 13,47 | 0,85 | 6,3% | 4,01 | 29,8% | 0,85 | 6,3% | 1,43 | 10,6% |
| C 18:0 | 4,37 | 0,25 | 5,8% | 0,98 | 22,3% | 0,25 | 5,8% | 0,47 | 10,8% |
| C 18:1 11t+10t | 0,74 | 0,05 | 6,5% | 0,16 | 22,0% | 0,05 | 6,5% | 0,08 | 10,8% |
| C 18:1 total | 11,20 | 0,80 | 7,1% | 2,76 | 24,6% | 0,80 | 7,2% | 1,47 | 13,1% |
| C 18:2 9c11t | 0,28 | 0,02 | 6,8% | 0,06 | 22,8% | 0,02 | 6,8% | 0,03 | 11,2% |
| C 18:2 total | 0,89 | 0,03 | 3,0% | 0,24 | 27,5% | 0,03 | 3,0% | 0,04 | 4,8% |
| C 18:3 total | 0,29 | 0,03 | 9,8% | 0,19 | 68,0% | 0,03 | 9,6% | 0,05 | 18,6% |
| SFA | 30,55 | 0,79 | 2,6% | 3,17 | 10,4% | 0,79 | 2,6% | 1,19 | 3,9% |
| MUFA | 12,74 | 0,84 | 6,6% | 2,87 | 22,5% | 0,84 | 6,6% | 1,53 | 12,0% |
| PUFA | 1,29 | 0,05 | 3,7% | 0,21 | 15,8% | 0,05 | 3,7% | 0,08 | 6,1% |
| Omega3 | 0,36 | 0,03 | 8,9% | 0,20 | 54,2% | 0,03 | 8,8% | 0,06 | 16,1% |
| Omega6 | 0,85 | 0,03 | 3,6% | 0,41 | 47,8% | 0,03 | 3,6% | 0,06 | 7,3% |
| Fat | 45,03 | 0,41 | 0,9% | 1,38 | 3,1% | 0,41 | 0,9% | 0,73 | 1,6% |



FTMIR monitoring (octobre 2009 -> Déc.2010)

1- Traceability and centralized anchorage through control milk sample common for the analysers:

-> LC = Control milk -80°C

⇒ to determine correction

-> LT = Liquid milk with bronopol at 4°C

⇒ to check and secure correction

2- Afterward standardization of predictions by additive correction from the national database:

| Mois | 12/09 | | 01/10 | | 02/10 | | 03/10 | | 04/10 | | 05/10 | | 06/10 | | 07/10 | | 08/10 | | 09/10 | | 10/10 | | 11/10 | | 12/10 | |
|----------|-------|----|-------|----|-------|----|-------|----|-------|----|-------|----|-------|----|-------|----|-------|----|-------|----|-------|----|-------|----|-------|----|
| Appareil | 1q | 2q |
| 1 | LT | LT |
| 2 | LC | LC | LT | LT |
| 3 | LC | LC | LT | LT |
| 4 | LC | LC | LT | LT |
| 5 | LC | LC | LT | LT |
| 6 | LC | LC | LT | LT |
| 7 | LC | LC | LT | LT |
| 8 | LC | LC | LT | LT |
| 9 | LC | LC | LT | LT |
| 10 | LC | LC | LT | LT |
| 11 | LC | LC | LT | LT |
| 12 | LC | LC | LT | LT |
| ... | LC | LC | LT | LT |



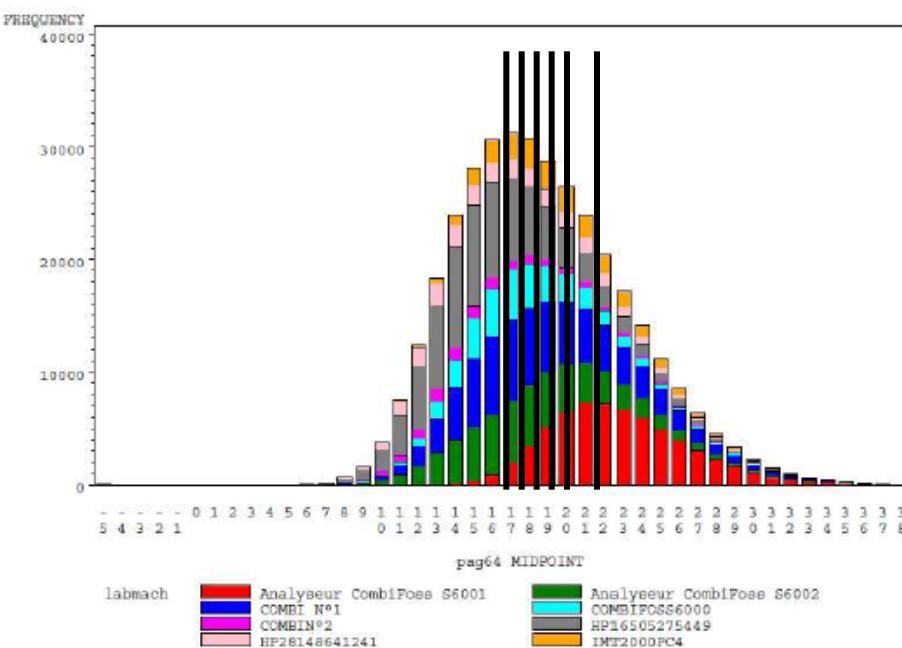
Un programme R&D pour les filières laitières de demain

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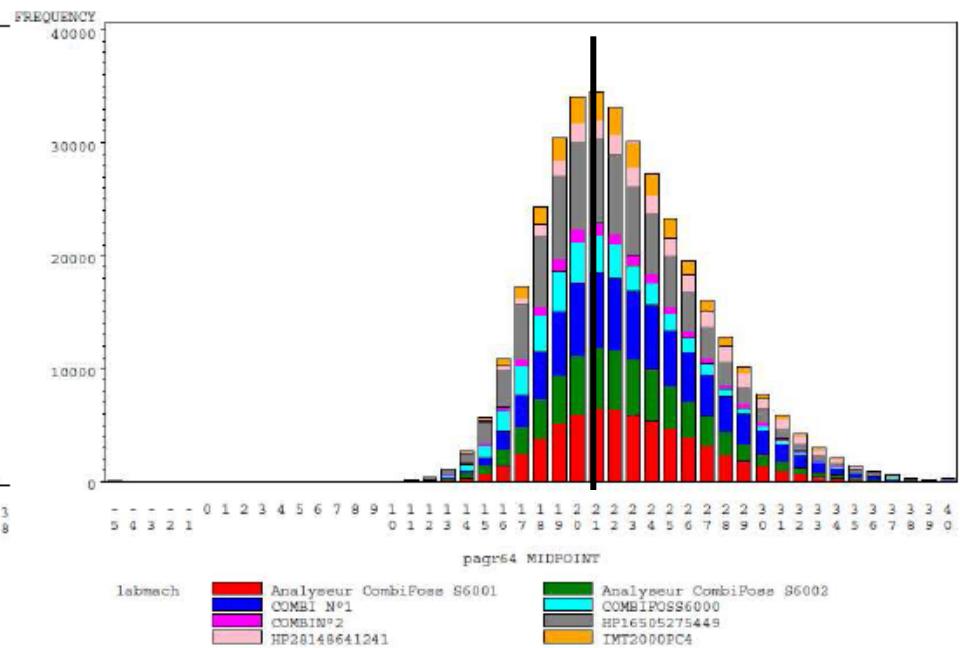
*Effect of data correction on a 14 month-long
population of results through a control milk network
8 FTMIR analysers*

Bovine milk – C18:1 %

Raw data



Corrected data





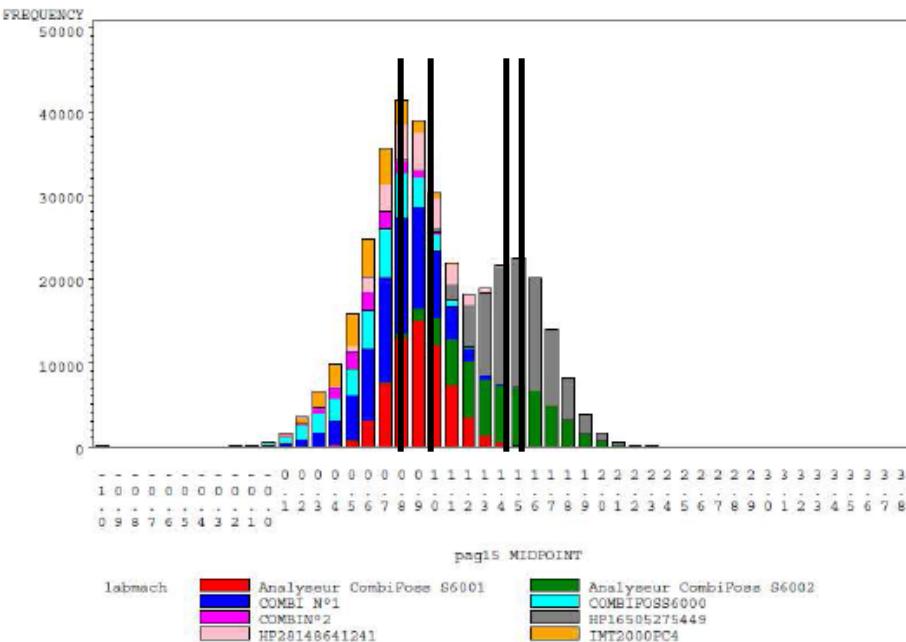
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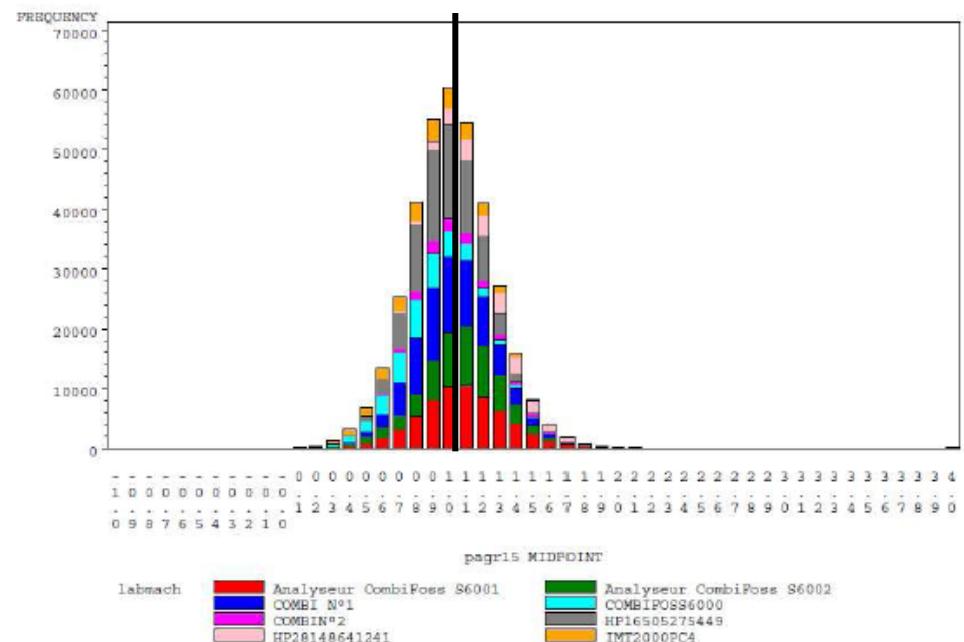
*Effect of data correction on a 14 month-long
population of results through a control milk network
8 FTMIR analysers*

Bovine milk – Omega3 %

Raw data



Corrected data





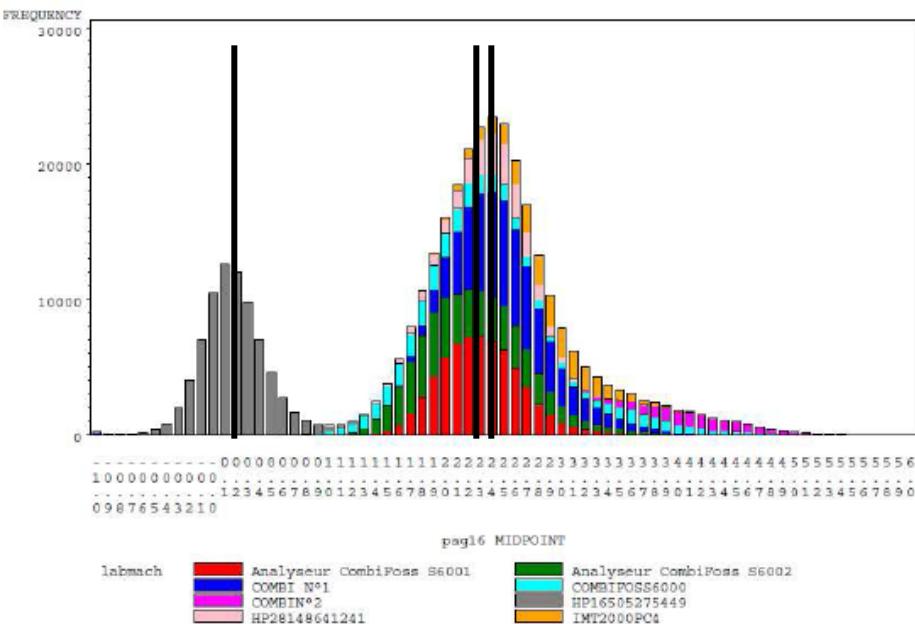
Un programme R&D pour les filières laitières de demain

PhénoFinlait

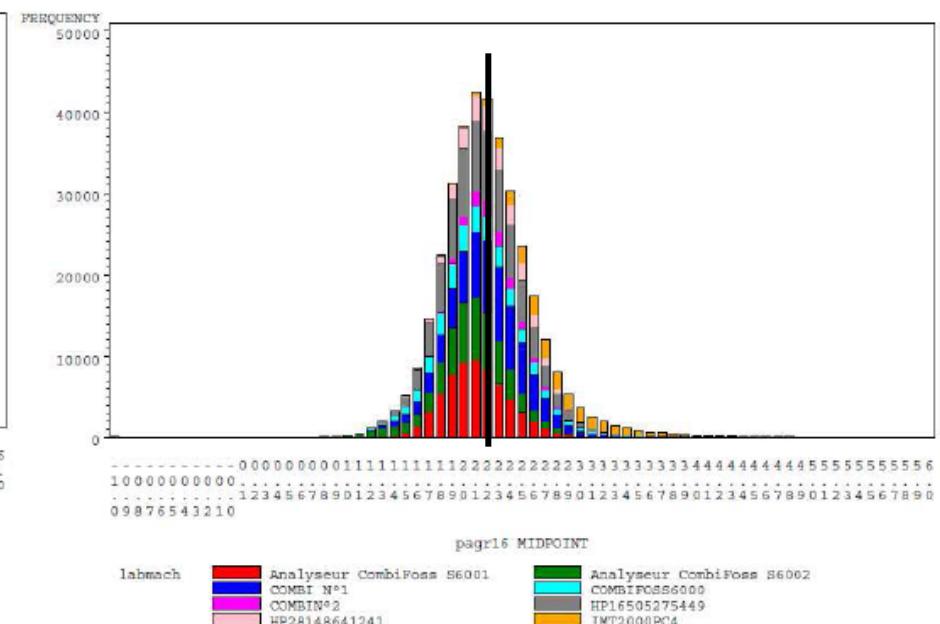
*Effect of data correction on a 14 month-long
population of results through a control milk network
8 FTMIR analysers*

Bovine milk – Omega6 %

Raw data



Corrected data





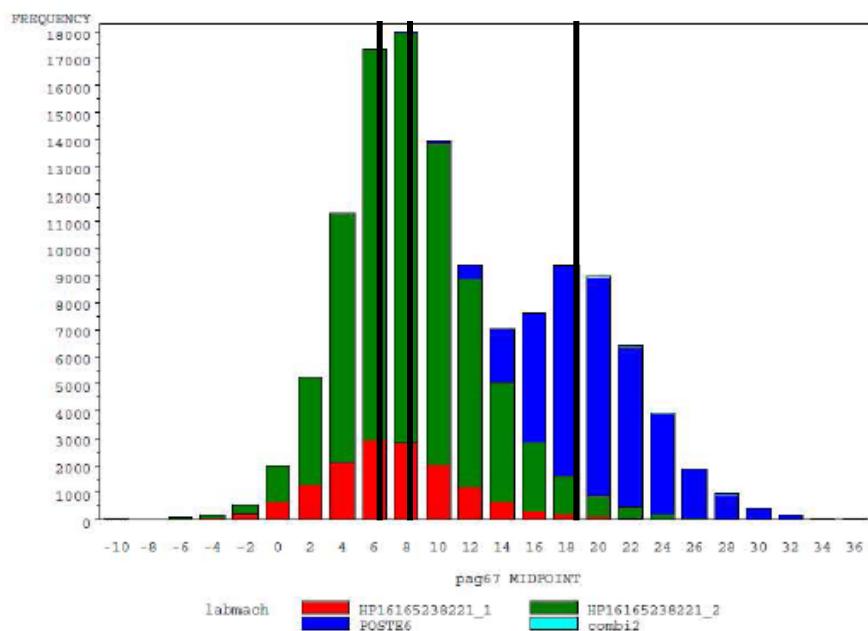
Un programme R&D pour les filières laitières de demain

PhénoFinlait

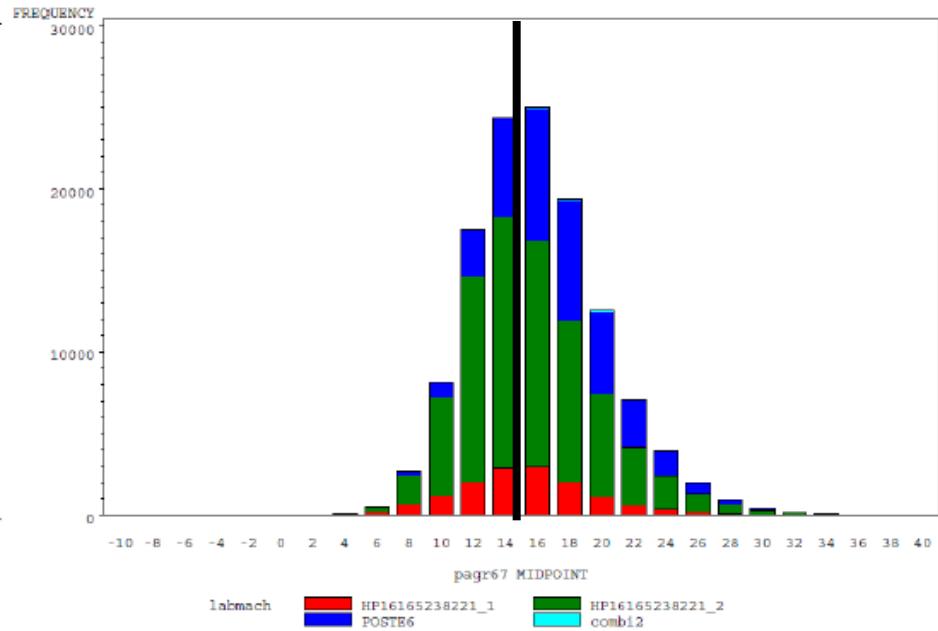
*Effect of data correction on a 14 month-long
population of results through a control milk network
4 FTMIR analysers*

Ovine milk – C18:1 %

Raw data



Corrected data





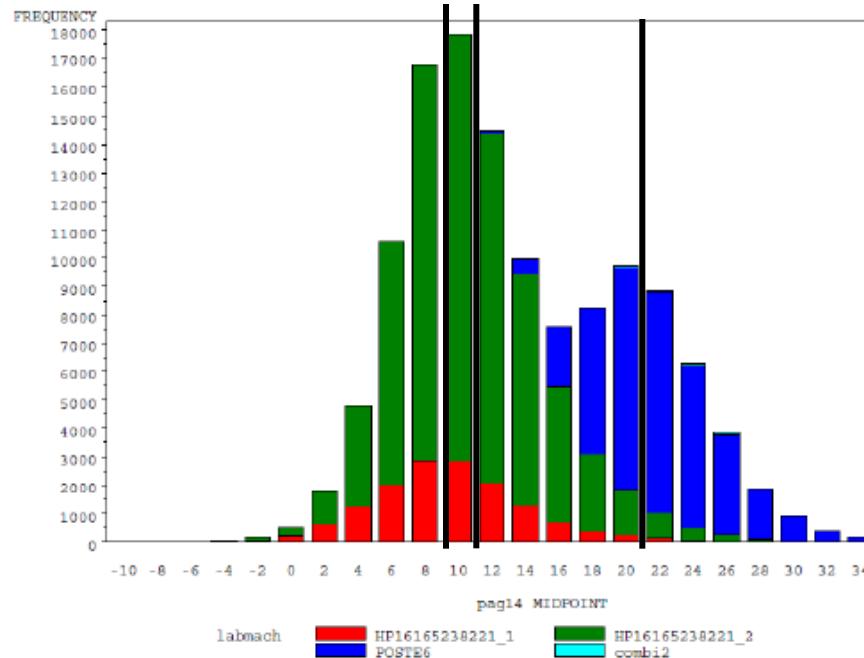
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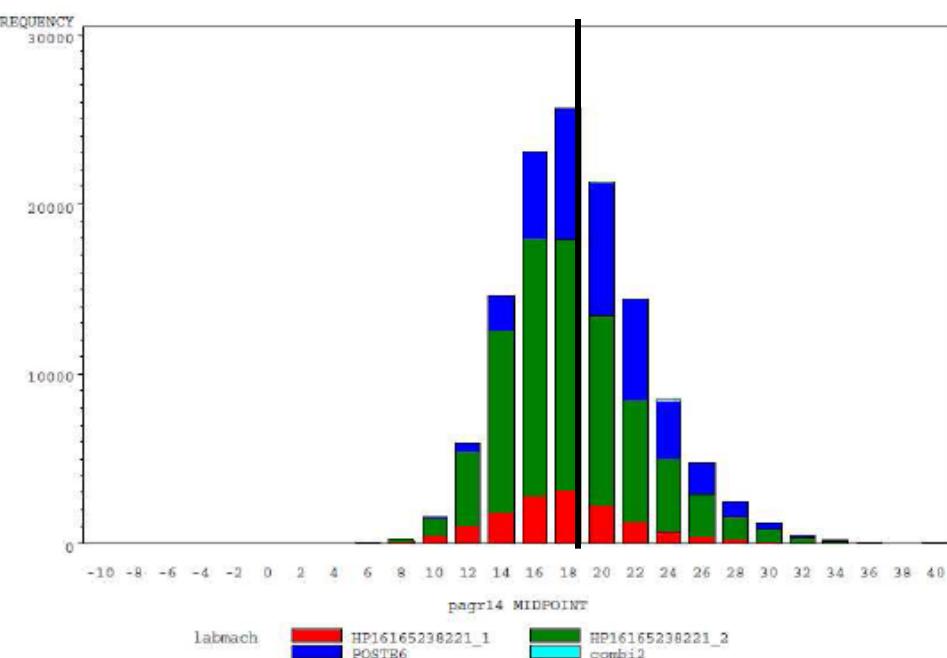
*Effect of data correction on a 14 month-long
population of results through a control milk network
4 FTMIR analysers*

Ovine milk – MUFA %

Raw data



Corrected data





Conclusion

- Spectrum standardization as applied did not suffice to harmonise results between instruments
- Efficient result standardization is achieved through centralised calibration thus providing good analytical characteristics r, R and R/r
- Standardisation can be achieved either at the level of predicted FA content values or at the level of absorbances
- Building a multi lab/instrument spectrum data base for multi purpose implies either collecting well standardised spectral data or include the means for afterward correction to minimise possible strong background noise and develop more efficient diagnosis tools.

Acknowledgment is given

to the companies Foss and Bentley which provided standard spectrum export facilities to laboratories,

to the partners of the programme, laboratories and DHI organisations, which provided data,

to the members of the scientific committee of Phénofinlait who advised for and managed that work.

Thank you for your attention